

Analytical methods

Determination of protease activity, AU(A) – analysis by Konelab

Scope

Novozymes Enzyme QC laboratories involved in analysis of samples from Novozymes production.

Application

The method is used to determine the protease activity in samples.

Principle

Dimethyl casein (DMC) is hydrolyzed by the proteolytic enzyme to small peptides as shown below:

The primary amino groups formed in this process react with trinitrobenzene sulfonic acid (TNBS), forming a colored complex as shown below:

$$O_2$$
N OH O_2

This color development is monitored by the Konelab, so the change in absorption per time unit can be calculated. This increase in absorption is proportional to the reaction rate, and thus to the enzyme activity.

Reaction conditions		
Temperature	50°C	
pH	8.3	
Substrate concentration	0.25%	
Enzyme concentration	0.006-0.017 mAU(A)/ml	
Reaction time	8 minutes	
Measuring time	2 minutes (seven measurements with an interval of 18 seconds)	
Wavelength	405 nm	

Definition of unit

The activity is determined relative to a protease A standard. The result is given in the same units as the standard, which is designated:

 $AU(A) - (\underline{A}nson \underline{U}nit (\underline{A} standard).$

Method parameters

Specificity

Serine endopeptidases with activity at pH 8.3 will be measured. Primary amines will interfere.

Range

The range is 0.072–0.216 mAU(A)/ml in the final dilution.

Limit of determination

The limit of determination is 0.001 AU(A)/g (for a minimum preparation of 1 g liquid sample in 10 ml).

The limit of determination is 0.002 AU(A)/g (for a minimum preparation of 1 g solid sample in 25 ml).

Accuracy and intermediate precision:

The accuracy is 99%.

The intermediate precision is 4%.

Equipment

-4b		
Equipment		
Konelab 30 Analyzer	Thermo Fisher Scientific NOTE: Critical equipment	
Diluter	e.g., Hamilton	
Analytical balance	e.g., Sartorius, Mettler	
Balance	e.g., Sartorius	
pH meter	e.g., Radiometer, Metrohm	
Thermometer (0–100°C)	-	
Magnetic stirrer plates		

Chemicals

Name	Chemical formula	Brand	Working environment guideline
Brij 35 (30% w/v)	-	e.g., Sigma B4184	Harmful
N,N-dimethyl casein (DMC)	-		-
Sodium tetraborate	Na ₂ B ₄ O ₇ , 10	e.g., Merck art. no. 6308	Avoid contact with
decahydrate	H ₂ O		eyes and skin
Sodium dihydrogen phos-	NaH ₂ PO ₄ ,	e.g., Merck art. no. 6346	-
phate monohydrate	H ₂ O		
Sodium sulfite	Na ₂ SO ₃	e.g., Merck art. no. 6657	-
2,4,6-trinitrobenzene	C ₆ H ₃ N ₃ O ₉ S	e.g., Fluka BioChemica art	Toxic, explosive in dry
sulfonic acid (TNBS)		no. 92822 1 M solution	form, corrosive

NOTE: Always check out the Material Safety Data Sheet (MSDS) for the chemicals marked with an environment guideline.

Reagents/substrates

Preparation of reagents and substrates must be documented according to valid procedures.

Brij 35 solution 15% w/v

EXAMPLE: Preparation of 2,000 ml Brij 35 15%:

Step	Action
1	Transfer 1,000 ml Brij 35 30% w/v to a 2,000-ml measuring flask. Brij 35 30% can be heated to 35–45°C
2	Rinse the Brij flask with demineralized water and transfer the water to the 2,000-ml flask
3	Fill up to 2,000 ml with demineralized water and mix vigorously
4	Brij 35 solution can be stored for 2 months in a refrigerator

TNBS, 2,4,6-trinitrobenzene sulfonic acid solution 0.1% w/v

EXAMPLE: Preparation of 100 ml TNBS 0.1%:

Step	Action
1	Pipette 0.345 ml 1 M TNBS into a 100-ml measuring flask
2	Fill up to 100 ml with demineralized water and mix
3	Measure pH. pH is approx. 2.5 ± 0.3 in a TNBS solution 0.1%
4	TNBS solution must be prepared every day.
	NOTE: The TNBS solution is stored in a refrigerator, shielded against light (brown bottle or aluminum foil)

IMPORTANT: In dry form, TNBS is classified as an explosive, and it is very important to prevent TNBS from drying out. Rinse empty TNBS bottles with water before disposal. Keep waste containers with cuvettes closed in order to prevent the TNBS in these from drying out. Do not store these waste containers for too long.

DMC, N,N-dimethyl casein solution 0.32%

EXAMPLE: Preparation of 1,000 ml DMC solution 0.32%:

Step	Action
1	Dissolve 3.20 g N,N-dimethyl casein (DMC) in approx. 200 ml boiling demineralized
	water in a beaker.
	Keep the beaker with the DMC on the heat while stirring for 20 minutes. Keep the
	temperature close to boiling point
2	Dissolve 25.92 g Na ₂ B ₄ O ₇ , 10 H ₂ O, and 13.30 g NaH ₂ PO ₄ , 1 H ₂ O in 400 ml demineral-
	ized water in a 1,000-ml measuring flask
3	After cooling, transfer the DMC solution to the 1,000-ml measuring flask
4	Fill up to 1,000 ml with demineralized water
5	Mix well and filter the solution twice through a Whatman filter no. 54 or a filter with
	similar pore size
6	Add 1.2 ml Brij 35 solution 15% and mix. Brij 35 is added after filtration to avoid for-
	mation of foam during filtration
7	Check the pH of the solution; pH must be 8.00 ± 0.05 . Adjust if necessary with HCl or
	NaOH
8	Fill the substrate into smaller vessels and store for 24 hours in a refrigerator. Then
	freeze the substrate
9	The DMC can be stored for 1 year in a freezer and for 1 week in a refrigerator after
	thawing.
	IMPORTANT: When removing DMC solution 0.32% from a freezer, stir the solution
	after thawing until it is clear

Sodium sulfite stock solution 20% w/v

EXAMPLE: Preparation of 10 L sodium sulfite stock solution 20% w/v:

Step	Action
1	Dissolve 1,000 g Na ₂ SO ₃ by vigorous agitation in a beaker that has first been filled with approx. 3.5–4 L demineralized water. Prepare this solution twice. IMPORTANT: Sprinkle Na ₂ SO ₃ carefully into the water in the beaker while stirring
2	Stir until the sodium sulfite in the beakers has dissolved
3	Transfer the sodium sulfite solution in the two beakers to a 10-L measuring flask
4	Fill up to 10 L with demineralized water and mix
5	The stock solution can be stored at room temperature for 1 month in a closed vessel

Sodium sulfite solution 2%

EXAMPLE: Preparation of 10 L sodium sulfite solution 2%:

Step	Action
1	Measure out 1,000 ml sodium sulfite stock solution 20% and transfer it into a 10-L measuring flask containing approx. 8 L demineralized water
2	Add 15 ml Brij 35 15%
3	Fill up to 10 L with demineralized water and mix
4	Sodium sulfite solution 2% can be stored for 2 days at room temperature

Handling of enzymes and chemicals

Enzymes and enzyme solutions should be handled in a fume hood or in closed containers. Avoid inappropriate handling of enzymes and enzyme solutions, which may result in aerosol/dust generation.

Avoid inhalation of dust aerosols and contact with skin and eyes.

Handling of chemicals and disposals of waste must be performed according to the MSDS for the product.

Standard

Protease A standard is available upon request.

Stock solution

Step	Action
1	Weigh out 1.08 AU-A of the protease A standard
2	Dissolve the standard in sodium sulfite solution 2% in a 500-ml measuring flask
3	Stir on a magnetic stirrer for 15 minutes
4	NOTE: The stability is set to 7 hours

Enzyme standards

Step	Action					
1	sulfite sol	ution 2% on curve with a	the diluter direct	ed by diluting the stoc ly into the sample cup lowest and highest st	s. The standard cur	rve
	No.	Dilution factor	Enzyme stock solution µl	Diluent sodium sulfite 2% µl	Activity mAU(A)/ml	
	1	30	40	1,160	0.072	
	2	24	50	1,150	0.090	
	3	20	60	1,140	0.108	
	4	16	70	1,130	0.135	
	5	15	80	1,120	0.144	
	6	12	100	1,100	0.180	
	7	10	120	1,080	0.216	
	NOTE: Sta	ndard final s	olutions in sampl	e cups are stable for 3	o min after dilution	n

QC sample

The preparation of the QC sample must be documented according to valid procedures.

QC sample

QC sample for AU(A) is available upon request.

Step	Action
1	Weigh out 2.11 AU(A) of the QC sample
2	Dissolve the QC sample in sodium sulfite solution 2% in a 250-ml measuring flask
3	Stir on the magnetic stirrer for 15 minutes
4	Dilute the stock solution 60 times with sodium sulfite solution 2% on the diluter.
	IMPORTANT: The final solution in the sample cup is stable for 30 minutes after dilution

Samples

The lowest dilution of liquid samples: 10 times. The lowest dilution of solid samples: 25 times.

Weighing and dilution of protease samples are described in the following:

Step	Action
1	Weigh out 0.5–1.5 g of sample
2	Dilute the sample in a measuring flask with sodium sulfite solution 2%
3	Stir for approx. 15 minutes on a magnetic stirrer
4	The samples are further diluted with sodium sulfite solution 2% on the diluter. The dilution on the diluter is made directly into the sample cup. NOTE: If possible, the activity in the final dilution should be approx. 0.144 mAU(A)/ml)
5	Each weighing is analyzed once on the Konelab. IMPORTANT: Sample final solutions in sample cups are stable for 30 minutes after dilution

Blank

Sodium sulfite solution 2% is used as a blank sample.

Procedure

Step			Ac	tion		
1	Prepare all samples.					
2	Place the reag	ents in the Ko	onelab:			
	Reagent	Konelab name	Reagent container volume	Syringe speed	Stability on board the Konelab	
	DMC 0.32%	DMC	60	Normal	1 day	
	TNBS solu- tion 0.1%	TNBS	20	Normal	4 hr	
3			sample, and sa	•	Konelab in the sta	ted order.
4	Start and run		,	-		

Configurations

The Konelab reagent and test definition for AU(A) is shown in appendix 1.

Calculation

drawn with the activities of the standards in mAU(A)/ml as the x-values and the A/min of the standards as the y-values. A logit/log4 algorithm is used The enzyme activity of the diluted samples is read from the standard curve. Calculation of activity of a sample in AU(A)/g is performed as stated in the formula: $Activity AU(A)/g = \frac{S \cdot V \cdot F}{W \cdot 1000}$ $S = \text{Reading from the standard curve in mAU(A)/ml}$ $V = \text{Volume of the measuring flask in ml}$ $F = \text{Dilution factor for second dilution}$ $W = \text{Weight of sample in g}$ $1000 = \text{Conversion factor from mAU(A) to AU(A)}$ $EXAMPLE: 0.5416 \text{ g sample is dissolved in a 250-ml measuring flask and further diluted 30 times using a diluter.}$ On the Konelab an A/min of 0.2224 is measured.	Action
drawn with the activities of the standards in mAU(A)/ml as the x-values and the A/min of the standards as the y-values. A logit/log4 algorithm is used The enzyme activity of the diluted samples is read from the standard curve. Calculation of activity of a sample in AU(A)/g is performed as stated in the formula: $ActivityAU(A)/g = \frac{S\cdot V\cdot F}{W\cdot 1000}$ $S = \text{Reading from the standard curve in mAU(A)/ml}$ $V = \text{Volume of the measuring flask in ml}$ $F = \text{Dilution factor for second dilution}$ $W = \text{Weight of sample in g}$ $1000 = \text{Conversion factor from mAU(A) to AU(A)}$ $EXAMPLE: 0.5416 \text{ g sample is dissolved in a 250-ml measuring flask and further diluted 30 times using a diluter.}$ On the Konelab an A/min of 0.2224 is measured.	The activity of the enzyme samples is determined relative to the standard curve
tion of activity of a sample in AU(A)/g is performed as stated in the formula:	
$ACTIVITY = {0.5416 \cdot 1000} = 1.68 \text{ AU(A)/g}$	The enzyme activity of the diluted samples is read from the standard curve. Calculation of activity of a sample in AU(A)/g is performed as stated in the formula:

Approval

Standard curve:

Parameter	Requirement
Curve appearance	The standard curve must be a smooth hollow curve. The rates for the standards must be at the level that is normal for the protease A standard curve using the relevant DMC and TNBS batches
r ²	\geq 0.995. If r^2 is < 0.995, one standard may be removed

Approval of QC sample:

Parameter	Requirement
QC sample	The result of the QC sample must not deviate from the control limits, which are set to: Declared value \pm 8%.

Approval of samples:

Parameter	Requirement
Sample	CV of six weighings (= three weighings on two different standard
	curves) ≤ 7.2.

Statement of analysis results

The analysis result is stated with three significant digits.

Configurations

AU(A) test definition:

Test definition	Konelah	Arena 6.5	Page:	
AU-A		nes S/N: S14 19 151 Kemisk Laboratorium		
Date : 2006-09-1 Time : 15.52	3	· • • • • • • • • • • • • • • • • • • •		
Full name Online Name Test type Result unit Number of Decim.		Test In Use Y Test limit 0 Initial absorbance 0 Dilution limit * Secondary dil 1+ 0 Critical limit * Reflex test limit *		Abs/min A Abs/min Abs/min Abs/min
Acceptance Dilution 1+	Automatic 0.0	Reference class L	OW HIGH	In Use
Sample type	Sample typ	ce 5 Correction factor Correction bias 0 Temperature 5	1.00 0.00 Abs/min	n
Calibration type Factor	None 1.00	Bias	0.00	
Bias correction in	use NO			
Cd reduction	NO			
Manual QC in Use	NO	Routine QC in Use	NO	
Blank	None	Fixed cuvette		
Reagent Disp. with Wash reagent Reagent wash	DMC Extra DMC Before dis	Volume (ul) Add. Volume (ul) spense	180 50	
Incubation		Time (sec)	480	
Reagent Disp. with Wash reagent Reagent wash	TNBS Extra TNBS Before dis	Volume (ul) Add. Volume (ul) spense	36 50	
Incubation		Time (sec)	60	
Sample Disp. with Dilution with	Extra Water	Volume (ul) Add. Volume (ul) Wash reagent	18 50 None	
Incubation		Time (sec)	360	
Measurement Wavelength (nm) Curve type Nonlinearity limit	s	Side wavel. (nm)	None	
Conc. (Abs/min) Time (sec)	0.00000	Percent (%) Points and Intervals	25 7 / 18.0	0 (sec)

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Reagent definitions

DMC:

Reagent definition	Konelab Are	na 6.5	Page: 1
	Novozymes Enzym Kemis		
13.09.2006 15:52			
Reagent	DMC	Lot	Expiry date (dd.mm.yy)
Stable on board (days) Alarm limit (ml)	1 2.0		
Information			
Vial volume Barcode id	60 ml		
Syringe speed	Normal		

TNBS:

Reagent definition	Konelab Ar	ena 6.5	Page: 1
		S/N: S14 1 sk Laborator	
13.09.2006 15:52			
Reagent	TNBS	Lot	Expiry date (dd.mm.yy)
Stable on board (days) Alarm limit (ml)	1		
Information			
Vial volume Barcode id	20 ml		

Validity

Valid from May 2010.

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